

Mouse IL-22 Precoated ELISA Kit

Cat#:1212202

[Product Description]

The Mouse IL-22 Precoated ELISA Kit is used for the in vitro quantitative detection of mouse IL-22 in mouse serum, plasma, buffer or cell culture medium. It can detect both natural and recombinant mouse IL-22. This kit is only used for scientific research. It shall not be used for diagnosis. Read the instruction and check the components of the kit before use. If there are any questions, please contact with Shenzhen Dakewe Bio-engineering Co., Ltd.

Detection range: 200~3.13 pg/mL

Sensitivity: 1 pg/mL

Precision: Intra-Assay CV: ≤10%; Inter-Assay CV: ≤15%

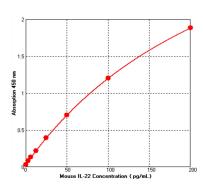


Figure 1.Note: Mouse IL-22 standard curve range: 200~3.13 pg/mL

[Materials Provided]

Product Name	Cat No.	Component Name	Size	Quantity	Reconstitution
	1212202				Lyophilized, diluted
		Cytokine Standard	48 T	2 vials	according to the
					instructions on the vial
		Biotinylated Antibody	50 μL	2 vials	1:100 diluted with
					Dilution Buffer R (1×)
		Streptavidin-HRP	50 μL	2 vials	1:100 diluted with
Mouse IL-22 Precoated ELISA Kit					Dilution Buffer R (1×)
		Dilution Buffer $R(1\times)$	10 mL	3 bottles	Ready-to-use
		Washing Buffer(50×) 15	15 mL	1 bottle	1:50 diluted with
					distilled water
		TMB	10 mL	1 bottle	Ready-to-use
		Stop Solution	10 mL	1 bottle	Ready-to-use
		Precoated 96-well strip	96 T	1 plate	Doody to use
		ELISA plates			Ready-to-use
		Plate Sealers	/	4 strips	Ready-to-use

[Self-provided Items]

- 1. Microtiter plate reader fitted with appropriate filters (450 nm required with optional 620 nm reference filter).
- 2. Adjustable Pipettes and pipette tips.
- 3. Distilled or deionized water, brand new filter paper.
- 4. Vortex mixer and magnetic stirrer.

[Cautions]

- 1. When the kit is not in use, it shall be stored according to the label on the vial or bottle. Pre-warmed to room temperature (18°C~25°C) before use. The lyophilized standards shall be discarded after use.
- 2. Streptavidin-HRP and Biotinylated Antibody shall be centrifuged briefly before use.
- **3.** The plate strips not used in the experiment shall be immediately put back into the package and sealed to prevent contamination. The reagents shall be packaged or covered when not in use.
- 4. When adding reagents, standards, and samples in experimental operations, please use disposable

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tips to avoid cross-contamination.

- 5. The solution or reagent is mixed evenly by shaking or stirring.
- 6. The order of adding reagents to the wells of the experimental plate shall be consistent to ensure the same incubation time for all reaction wells.
- 7. Clean plastic containers are used for washing liquid.
- 8. In the process of washing, the washing liquid is removed by suction or shaking off, and then fully patted dry on the filter paper. Do not put the filter paper directly into the reaction well to absorb water.
- 9. TMB is sensitive to light, so long time exposure to light shall be avoided, and metal contact shall be prevented from affecting the results. If the TMB to be used turns blue, it indicates that the TMB has been contaminated, so please discard it. Please read the OD value within 10 minutes after stopping the reaction.
- 10. Please incubate strictly according to the time and temperature indicated in the instructions.
- 11. Do not use expired kits, and do not mix reagents from different lots.
- 12. Do not exceed the linear range. The results beyond the linear range are non-linear, and accurate results cannot be obtained according to this standard curve.
- **13.** Specificity: Does not react with other mouse cytokines.

[Preparation of Reagents]

1. Cytokine Standard: The lyophilized powder is resuspended according to the label instructions; let it stand for 5 minutes, and shake it slightly until it is completely dissolved. Then it is diluted with Dilution Buffer R(1×) to 200 pg/mL \, 100 pg/mL \, 50 pg/mL \, 25 pg/mL \, 12.5pg/mL \, 6.25pg/mL, 3.13 pg/mL.

Dilution step: Dissolve the Cytokine Standard according to the bottle label and transfer it into an EP tube, labeled as C7. Take another 6 EP tubes, labeled as C6/C5/C4/C3/C2/C1, and add 250 μL of Dilution Buffer R(1×) to each tube. Take 250 μL of Cytokine Standard from C7 and transfer it to C6 tube for 2-fold dilution. Dilute C5/C4/C3/C2/C1 tube in the same way. Only add Dilution Buffer $R(1\times)$ to C0 tube.

Standard code	Dilution ratio	Dilution Buffer R(1×) (µL)	Standard Concentration (pg/mL)
C7			200
C6	1: 2	250	100
C5	1: 4	250	50
C4	1: 8	250	25
C3	1: 16	250	12.5
C2	1: 32	250	6.25
C1	1: 64	250	3.13
C0		250	0

2. Biotinylated Antibody: By referring to the following table, it is diluted with Dilution Buffer R (1×) into a clean tube at the ratio of 1:100, and mixed well to make 1×working solution.

Number of Wells	Biotinylated Antibody (µL)	Dilution Buffer R(1×) (μL)
16	17	1700
24	25	2500
32	34	3400
48	50	5000
96	100	10000

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3. Streptavidin-HRP: By referring to the following table, it is diluted with Dilution Buffer R $(1\times)$ into a clean tube at the ratio of 1:100, and mixed well to make $1\times$ working solution.

Number of Wells	Streptavidin-HRP (µL)	Dilution Buffer R(1×) (µL)
16	17	1700
24	25	2500
32	34	3400
48	50	5000
96	100	10000

4. Washing Buffer $(50\times)$: It is diluted with distilled water at 1:50.

[Operation Process]

- 1. Before use, all reagents are mixed thoroughly to avoid foaming.
- 2. According to the number of experimental holes (blank and standard), the number of plate strips required is determined. Samples, standards and blanks shall be tested in duplicate well.
- 3. The 100 μ L of diluted Cytokine Standard is added to each standard well. 100 μ L of sample is added to each sample well. And 100 μ L of Dilution Buffer R (1×) is added to the blank control wells and incubated at 37 °C for 90 minutes.
- **4.** The liquid in the wells is removed; 300 μL of 1×Washing Buffer is added to each well, and the liquid is discarded after staying for 1 minute. Repeat 4 times. The wells are dried on the filter paper for the last time.
- **5.** The 100 μL of diluted Biotinylated Antibody is added to each well. Covered with the plate sealer and incubated at 37 °C for 60 minutes.
- 6. Repeat Step 4.
- 7. The 100 μL of diluted Streptavidin HRP is added to each well. Covered with the plate sealer and incubated at 37 °C, for 30 minutes.
- 8. Repeat Step 4.
- 9. The 100 μ L of TMB is added to each well. Incubated for 10~20 minutes in dark.
- 10. The $100~\mu L$ of Stop Solution is quickly added to the wells to stop the reaction. Within 10~minutes after stopping, the absorbance values are read at 450~nm. If the reader is capable of reading at 620~nm, the absorbance at 620~nm can be subtracted from the absorbance at 450~nm.

[Data Analysis]

- A). Logarithm of coordinate or natural number of fitting curve is recommended. The fitting equation is usually a straight line, a quadratic equation and a four-parameter equation. The best standard curve is selected by fitting of various application software, and the corresponding concentration is found according to the OD value of the sample.
- B). The diluted sample shall be multiplied by the dilution multiple when calculating the concentration.

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[List of Operation Steps]

 $100 \ \mu L$ of sample and standard are added to each well, and the control is set

Incubated at 37°C for 90 minutes and washed for 4 times

100 μL of diluted Biotinylated Antibody is added to each well $^{\sqcap}$

Incubated at 37°C for 60 minutes and washed for 4 times

100 μL of diluted Streptavidin HRP is added to each well

Incubated at 37°C for 30 minutes and washed for 4 times \Box

100 μL of ready-to-use TMB is added to each well and incubated in the dark place for 10~20 minutes

100 μL of Stop Solution is added to each well $\hfill \Pi$

The plate is read at 450 nm in 10 minutes

[Storage]

The kit is stored at 2°C~8°C and can be stored stably for 12 months. The kit cannot be used after expiration.

[Appendix]

Troubleshooting Guide			
Problems	Possible reasons	Solutions	
	(1) Incorrect washing method;	(1) The correct method is used to wash the strips	
	(2) Reagents have expired	(2) Do not use the expired reagents	
1. Very weak results	(3) Enzyme conjugate diluent concentration is too low	(3) Repeat the experiment according to the recommended dilution concentration of the manufacturer's instructions	
	(4) Inefficient incubation time	(4) Repeat the test according to the incubation time of each step	
	(5) Incorrect storage of reagents	(5) Reagents are stored correctly to avoid repeated freezing and thawing	
	(6) Wrong filter in microplate reader was used	(6) The correct microplate reader wavelength setting is used	
The repeatability of standard curve and determination is poor	(1) Incorrect washing method	(1) The correct method is used to wash the strips	
	(2) Uneven dilution	(2) Return the reagent to room temperature, and the sample and reagent are gently diluted	
	(3) Plates are not clean	(3) The bottom of the plate is wiped clean before detecting the wavelength	
	(4) Wrong filter in microplate	(4) The correct wavelength parameters	

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		Research Use On	
	reader was used	of microplate reader are set up	
	(5) Reagents from different lots	(5) The reagents in the same batch of	
	are mixed and cross-used	kits are used	
	(6) Cross contamination in the		
	well	(6) Disposable suction tips are used to	
	Inconsistency of incubation time,	add sample or standard to avoid cross	
	washing plate and color	contamination	
	development time		
	(7) Samples and reagents are	(7) The liquid is released near the	
	added to the non-coated area	middle of the plate well	
	(8) Incorrect washing, incubation	(8) Time was used to correct the	
	and color development time	incubation time of each step	
	(1) There are problems in the	1	
	preparation of the plate washer, for	(0) P	
3.	example, the measuring cylinder is	(9) Repeat test, and make sure that the	
Blank plate	not clean, containing enzyme	reagent does not contain enzyme	
(positive	inhibitors (such as sodium azide),	inhibitors	
control does	etc		
not develop	(2) Wrong reagents were added or	(10) Read the instructions carefully and	
color)	omitted	repeat the test with the correct reagents	
	(3) Reagents have expired	(11) Do not use the expired reagents	
	(1) Incorrect washing method	(12) The correct method is used to wash	
		the strips	
	(2) The color developing fluid has	(13)The color developing solution	
	deteriorated	before use shall be colorless	
	(3) Reagents have expired	(14) Do not use the expired reagents	
	(4) Incorrect reagent dilution, for	(15) Repeat the experiment according to	
4.	example the concentration of	the recommended dilution concentration	
Blank	enzyme added is too high	of the manufacturer	
background is	(5) Distilled water is contaminated		
high	by enzymes	(16)Fresh distilled water is used	
		(17) When repeating the test, i.e. adding	
	(6) Contaminated by other positive	samples and washing, be careful not to	
	samples	cross-contamination	
	(7)The temperature of the	(18) The color reaction time is appropriately shortened	
	incubator exceeds 37°C or the		
	reaction time is too long		

[Company Information]

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